

Contribution of Legumes on Phosphoric Absorption by *Panicum maximum* cv Riversdale in Intercropping System

Sajimin¹, Purwantari ND¹, Sugoro I²

¹Indonesian Research Institute of Animal Production, Ciawi PO Box 221, Bogor Indonesia

²PATIR BATAN, Jakarta Indonesia

E-mail: djiemin@yahoo.com

(received 24-06-2016; revised 23-08-2016; accepted 29-08-2016)

ABSTRAK

Sajimin, Purwantari ND, Sugoro I. 2016. Kontribusi tanaman legume dalam peningkatan serapan fosfor rumput *Panicum maximum* cv Riversdale dengan sistem tanam tumpang sari. JITV 21(3): 151-158. DOI: <http://dx.doi.org/10.14334/jitv.v21i3.1520>

Ketersediaan fosfor (P) dalam tanah sebagai unsur hara yang sifatnya mobil mempengaruhi pertumbuhan tanaman. Tujuan dari penelitian ini untuk meningkatkan ketersediaan fosfor dan produksi rumput *Panicum maximum* cv Riversdale yang ditanam tumpang sari dengan tanaman leguminosa. Percobaan dirancang secara acak lengkap dengan lima perlakuan dan lima ulangan yaitu : A. *Gliricidia sepium* + *P. maximum*; B. *Calliandra calothyrsus* + *P. maximum*; C. *Leucaena leucocephala* cv Taramba + *P. maximum*; D. *Calopogonium mucunoides* + *P. maximum*; E. *P. maximum* (kontrol negatif). Tanaman tumpang sari ditumbuhkan pada pot dengan diberi pembatas yang dilubangi untuk pertumbuhan akar tanaman leguminosa di media rumput. Setelah tiga bulan tumbuh pada area tanaman leguminosa isotop ³²P diinjeksi sebanyak 50 ml (11,23 µci/ml). Setelah inkubasi selama 21 hari sampel tanah pada kedua area dan kedua tanaman dikumpulkan untuk analisis kadar isotop pada tanaman dan translokasi fosfor dideteksi dengan menggunakan perunut isotop ³²P. Produksi hijauan juga diamati pada ke dua tanaman. Hasil penelitian menunjukkan kadar fosfor tanah rumput dipengaruhi jenis legum, *G. sepium* dan *C. calothyrsus* nyata lebih tinggi terhadap kontrol, sedangkan *L. leucocephala* tidak berbeda nyata, dan kadar P nyata lebih rendah pada *C. mucunoides*. Deteksi ³²P menunjukkan akar legum yang terintegrasi di daerah rumput memindahkan P. Produksi rumput *P. maximum* dengan sistem tumpang sari secara nyata meningkat pada *G. sepium*, secara tidak nyata pada *L. leucocephala* dan *C. calothyrsus* dibandingkan perlakuan kontrol, sedangkan *C. mucunoides* turun 26,2% walaupun tidak berbeda nyata. Data dari tumpang sari *C. mucunoides* menunjukkan produksi hijauan legum tertinggi. Dapat disimpulkan bahwa ketersediaan fosfor dan produksi rumput dapat ditingkatkan dengan sistem tumpang sari dengan legum. Jenis legum mempengaruhi efektivitas.

Kata Kunci: Produksi Hijauan, Leguminosa, *Panicum maximum*, Tumpang Sari, Isotop ³²P

ABSTRACT

Sajimin, Purwantari ND, Sugoro I. 2016. Contribution of legumes on phosphoric absorption by *Panicum maximum* cv Riversdale in intercropping system. JITV 21(3): 151-158. DOI: <http://dx.doi.org/10.14334/jitv.v21i3.1520>

Phosphorus availability in soil as a mobile mineral influences forage growth. The purpose of doing this research is to enhance the soil phosphorus availability and grass production of *Panicum maximum* cv Riversdale by intercropping system with legumes. The experiment was conducted based on with randomized design with five treatments of mixcropping of: (i) *Gliricidia sepium* + *P. maximum*; (ii) *Calliandra calothyrsus* + *P. maximum*; (iii) *Leucaena leucocephala* cv Taramba + *P. maximum*; (iv) *Calopogonium mucunoides* + *P. maximum*; (v) *P. maximum* as negative control. Plants were grown in pots with split-root technique using partition with a whole to allow some legume roots grew in the grass side. After growing for three months, on the legume areas ³²P isotop solution was injected for 50 ml (11.23 µci/ml). After 21 days incubation samples were collected from both soil areas and both plants. The translocation of ³²P was determined using geiger counter from legumes into the grass and the concentration of ³²P was also determined in all plants. Forage productions was determined both in the legumes and grass. Result showed that soil phosphorus concentration in the grass area was significantly improved by intercropping with *G. sepium* and *C. calothyrsus*, while the one with *L. leucocephala* was similar, and the one with *C. mucunoides* was significantly lower than that of negative control (without legume). Detection of ³²P showed that there was P translocation in the system. *P. maximum* grass production depending on kind of legumes (P<0.05) those with *G. sepium* was significantly higher, *L. leucocephala* and *C. calothyrsus* were not significantly higher, while the one with *C. mucunoides* was 26.2% lower from the control although not significantly. However, *C. mucunoides* produced the highest forage from the legume plant. It is concluded that grass-legume intercropping had a positive impact on phosphorus soil concentration in the grass area and grass production. Kind of legumes influenced the effectivity.

Key Words: Forage Production, Legume, *Panicum maximum*, Intercropping, ³²P Isotop

INTRODUCTION

Developing forage plantations in the marginal areas such as acid areas is not optimal for crop production, but potentially important for feed forage. Horst et al. (2006) reported that acid soil reaches 1.7 billion hectares and 43% is in tropical area. In Indonesia, acid land reaches 102.8 - 107.4 million ha and has not been used optimally (Mulyani et al. 2004; Agus et al. 2015). Farmers in Indonesia used excessive phosphorus chemical fertilizer to enhance the crop production for 100 kg TSP/ha of paddy field. In the long term excessive P application might form cementing layer on land surface (Karama et al. 1991; Saidu & Abayomi 2015).

Phosphorus (P) is absorbed as orthophosphate ions is an important element for growing plant cells for phospholipid in the cell membrane, for accumulation and releasing cell energy in metabolism and as sugar-phosphate in nucleotides for genetic information (Franzini et al. 2009). P deficiency showed retarded growth in crops and reddish leaves due to the increase of anthocyanin. P is important for metabolism including cell division, respiration, and photosynthesis (Richardson et al. 2009). Hakim et al. (1986) suggests that in high acidity soil (pH <5), phosphate ions are easily binded with Al, Fe or Mn forming insoluble compounds that reduce the P availability. Together with N, P is important for forage quality and the availability are influenced by microbial activities surrounding the roots (Guo et al. 2000; Alan et al. 2009).

Good combination of intercropping or mixed cropping system of legumes and grass increases the grass production as well as reduce the use of N and P inorganic fertilizers or results friendly ecosystems (Exner et al. 1999). Intercropping of legumes and maize increases N in the soil (Li et al. 2003; Eskandori et al. 2009; Belel et al. 2014). The intercropping of legumes and elephant grass in the contour system also reduce the erosion and increase the production and forage quality (Anantawiroon et al. 2006; Mutegi et al. 2008).

Intercropping of cowpea-maize improves soil phosphorus availability and maize yields (Latati et al. 2014). Other researchers also reported that the non legume plants get phosphorus from legumes (Elgersma et al. 2000). Transportation of phosphorus from the legumes to the *Panicum* grass may be traced using radioisotope of ³²P injected in the legume areas. The intercropping system might be carried out using partition to separate both plants except that some of legume roots were in the grass area. The roots may

translocate the radioisotope from the legume areas. Therefore, the transportation of ³²P may be traced.

This research was aimed to increase the yield of the *Panicum maximum* (*Panicum* grass) in acid soil by intercropping with kinds of legumes incorporated with ³²P.

MATERIALS AND METHODS

Kinds of soils

This study was conducted in the greenhouse of IRIAP, Ciawi-Bogor using red-yellow podsolit soil collected from experimental garden. Soil was dried and sieved by 2 mm and then its nutrient content was analyzed. The results showed that soil consisted of 7% sand texture, 64% ash, and 29% clay at pH of 4.6. The organic and inorganic elements were 2.39% C, 0.097 % N, 61.67 ppm P, 0.08 ppm K, 5.72 ppm Ca, 1.09 ppm Mg and 0.31 ppm Na.

Design of experiment

This study employed completely randomized design with five treatments and five repetitions as follows:

- a) *Gliricidia sepium* + *Panicum maximum* cv Riversdale
- b) *Calliandra calothyrsus* + *P. maximum* cv Riversdale
- c) *Leucaena leucocephala* cv Taramba + *P. maximum* cv Riversdale
- d) *Calopogonium mucunoides* + *P. maximum* cv Riversdale
- e) *P. maximum* cv Riversdale only as negative control

If there were significant difference in analyses of variance, data were further analyzed with Duncan (Gomez & Gomez 1984).

Intercropping system

Intercropping system used in this study was Split-Root Technique (Catchpoole 1988) in combination with method by Xiao et al. (2004). One pol grass and one pol of each legume were planted in a pot containing 32 kg dry weight of soil and divided by a diagonally fiber partition. The partition had a hole so that the legume root partially passed through the hole in the area of the grass plant (Figure 1).

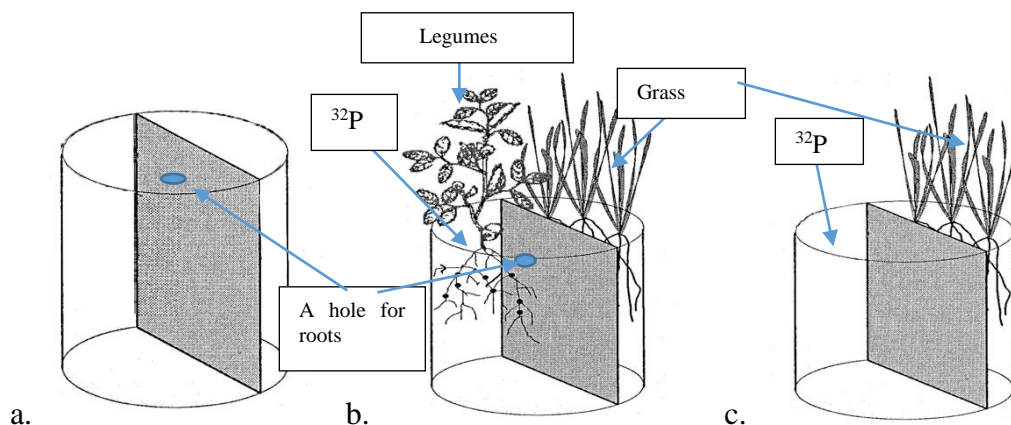


Figure 1. Position of the plants and ^{32}P injection in the pots with the fiber partition. (a) Pots before the plants growing; (b) Pots after the plants growing, some of legume roots grew through the hole to the grass area; (c) Control pots without legumes.

Application and detection of ^{32}P

Radioisotope ^{32}P as $\text{KH}_2^{32}\text{PO}_4$ solution was injected 50 mL (561.5 μCi /pots) in the soil area of legumes at three months growth. The transportation of the ^{32}P was traced using Geiger Muller all on the surface soil and part of the plants in IRIAP, while the concentration of ^{32}P was determined in the laboratory of National Nuclear Energy Agency of Indonesia. Distribution of the radio isotope in the stem pit and leaves of legumes was detected after the injection for 14 days. All data were collected for phosphorus recovery determination.

Total phosphorus content in the soil before and after the experiment was determined in Indonesian Center for Agricultural Land Resources Research and Development. Variables determined for forage production were plant height, number of shoots and bundles respectively for the legumes and the *Panicum* grass, dry weight of nodules, dry weight of roots, and forage production.

RESULTS AND DISCUSSIONS

Soil phosphorus availability. P availability of soils for grass or legumes was determined after the plants grew, but before P-isotope was introduced (Table 1). The soil planted only with *P. maximum* (control) at 64.0 ppm was lower than those integrated with legume roots of *G. sepium* (74.3 ppm) and *C. calothyrsus* (70.7 ppm), similar of *L. leucocephala* (65.7 ppm), but higher than the one with *C. mucunoides* (56.7 ppm). The P-availability in soils of *G. sepium* and *C. calothyrsus* were not significantly different. P-availability of soils planted with legume seemed higher than that of grass only (negative control). The P-availability in soils were influenced by legume intercropping and also kinds of legumes incorporated, the lower P in the legume area

affected the lower P in the grass area. The highest P was observed from *G. sepium* and *C. calothyrsus*, followed by *L. leucocephala* cv. *Tarramba*, and negative control. P in the grass area intercropped with *C. mucunoides* was significantly lower than that of the negative control.

The increase of P-availability in the grass area was in agreement with the one reported by Latati et al. (2014) that discover the increase of P in maize soil area integrated with cowpea. The lowest P in the legume soil area might be related with the P- consumption for the leave production. Leave production of *C. mucunoides* is relatively high. The lowest P-availability in the grass area integrated with *C. mucunoides* was not only affected by the low P in the legume area, but might be also influenced the availability of its root in the grass area. The effectiveness of P-transportation from legume soil area to the grass area was then detected by the transportation of ^{32}P isotop.

Recovery of ^{32}P on the legumes and grass

Data of ^{32}P tracers in each repetition were similar, therefore detection of ^{32}P using Geiger Muller counter was used to detect the P transportation from legumes soils to grasses's and part of both plants (Table 2). The isotop was transported from the legume soils in the whole parts of legumes and transported to grass soils and parts of grass. The transportation from legumes to grass areas was more influenced by kind of legumes than those from the concentration of ^{32}P in the soils like the one observed in *L. leucocephala*.

The injection of ^{32}P was carried out after the legumes producing roots in the grass areas. The ^{32}P was more detected in the legumes than that in grass except for *C. mucunoides*. The stem of *L. leucocephala* had the highest traced, while from other parts and other

legumes, as well as grass. The highest trace was observed in *G. sepium* dan *C. mucunoides*.

Data of tracers showed that ³²P *G. sepium* was higher in its stem than that in its leave tips. ³²P has more mobility in the stem than in the leaves. After pruning plants usually will grow again and the ³²P will be spread

out following the cell division (Kalaivanan et al. 2014). The Geiger Muller counter is more to detect the isotop transportation in the plant parts, therefore to observe the effect of the ³²P injection to the legumes and grass the concentration of ³²P of all plants were detected (Table 3).

Table 1. P availabilities on soils grown with legumes and on grass areas integrated with roots of legumes

Kinds of legume roots integrated with <i>P. maximum</i>	P on areas of legume soil (ppm)	P on areas of grass soils (ppm)
<i>G. sepium</i>	118.3 ^a	74.3 ^a
<i>L. leucocephala</i> cv. <i>Tarramba</i>	74.7 ^c	65.7 ^b
<i>C. calothyrsus</i>	107.3 ^b	70.8 ^a
<i>C. mucunoides</i>	78.7 ^c	56.7 ^c
Control	ND	64.0 ^b

Different superscript letters in the same column show significant difference (P<0.05); ND was not determined.

Table 2. Distribution of ³²P on legumes and grasses and their soil surface

Kind of legumes	Recovery of ³² P on legume soils and plants (µci/g.min)			
	Soil surface	Stem	Leaves	Leave tips
<i>G. sepium</i>	3391.7 ^{ab}	366.7 ^{ab}	216.7 ^a	158.3 ^a
<i>L. leucocephala</i>	8000.0 ^a	583.3 ^a	158.3 ^b	100.0 ^b
<i>C. calothyrsus</i>	1116.7 ^b	283.3 ^b	183.3 ^b	133.3 ^a
<i>C. mucunoides</i>	1250.0 ^b	350.0 ^{ab}	337.5 ^a	150.0 ^a
Control	2333.3 ^{ab}	ND	ND	ND
	Recovery of ³² P on grass soils and grass (µci/g.min)			
<i>G. sepium</i>	91.7 ^b	216.7 ^b	208.3 ^{ab}	233.3 ^a
<i>L. leucocephala</i>	91.7 ^b	258.3 ^b	250.0 ^{ab}	183.3 ^b
<i>C. calothyrsus</i>	166.7 ^a	275.0 ^b	166.7 ^b	175.0 ^b
<i>C. mucunoides</i>	150.0 ^a	350.0 ^a	300.0 ^a	275.0 ^a
Control	ND	ND	ND	ND

Different superscript letters in the same column show significant difference (P<0.05); ND was not determined. Traced of ³²P was carried out by Geiger Muller.

Table 3. Trace of ³²P (µci/g plants) on the legumes and grasses

Kind of legumes	Legumes	Grasses
<i>G. sepium</i>	839 ^d	11133 ^a
<i>L. leucocephala</i>	4945 ^b	9107 ^a
<i>C. calothyrsus</i>	2361 ^c	8043 ^a
<i>C. mucunoides</i>	14540 ^a	9673 ^a
Control	No plants	71 ^b

Different superscript letters in the same column show significant difference (P<0.05).

Each legume had significant difference of ^{32}P concentration. The highest concentration of ^{32}P on the legumes was observed at *C. mucunoides* followed by *L. leucocephala*, *C. calothyrsus*, while *G. sepium* had the lowest concentration (Table 3). Although no legume was grown in the control, ^{32}P was detected in the grass with very low concentration. This very low concentration resulted in none significant different at ^{32}P concentration in each grass intercropped, although there was 28 % different from the one integrated with *G. sepium* vs with *C. calothyrsus*. The highest ^{32}P concentration in the grass was observed at the one intercropped with *G. sepium* root (11133 $\mu\text{ci/g}$), followed by *C. mucunoides* (9673 $\mu\text{ci/g}$), *L. leucocephala* (9107 $\mu\text{ci/g}$) and *C. calothyrsus* (8043 $\mu\text{ci/g}$). Contribution of ^{32}P from the legumes into the grass depended on kinds of legumes. The grass integrated with *G. sepium* showed certain condition that the ^{32}P was low in the legume but it was high in the grass. The relation of the P concentration toward the grass production will be discussed in the grass production paragraph. Anantawiroon et al. (2006) reported that kinds of legumes in the intercropping system resulted in different production and quality of Napier grass.

Compared to the control the grass with legume root integration had higher ^{32}P . This result is in agreement with that reported by Richardson et al. (2009) that ^{32}P from legume areas is trans located to barley. The translocation is influenced by the legume root amount in the barley areas and legume morphology. The ^{32}P in

the grass areas influences the grass phosphoric absorption.

Root nodules and weights

Each legume function in nitrogen fixation had different root nodules in shapes, location in the roots and numbers (Table 4). Numbers of the nodules was expressed in weight, higher weight shows higher numbers. The heighest weight nodules were observed in *L. leucocephala*, however, they were only in the center (primary root). Therefore, the effectivity of the noodles only functioned for the legume not for the grass. Nodules of *G. sepium* spread over primary and secondary roots in high number including in grass area, therefore it would influence better for the grass production. The root nodules of *C. mucunoides* were quite a lot in the grass area, however, their quality was not as good. The color was black. The best quality of root nodules for nitrogen fixation is when they are pink and large.

Data of Table 5 shows that legume roots grew together with grass roots. This rhizofer system helps the translocation of nutrients including P from legume areas to grass area. The root nodules of the legumes especially those in the grass area will also help the nitrogen fixation for the grass growth. Rhizosfer zone in the grass areas will be influenced by the root legume structure (Fustec et al. 2010), while root activity significantly influences the physical, chemical and

Table 4. Root nodules of the legumes

Kind of legumes	Weight (g/plants)		Shape and colours	Position
	Legume areas	Grass areas		
<i>G. sepium</i>	1.38 ^b	0.45 ^b	Round, cream	Spread
<i>L. leucocephala</i>	3.94 ^a	0.06 ^c	Large, pink	Centre
<i>C. calothyrsus</i>	0.74 ^c	0.10 ^c	Spherical, pink	Centre
<i>C. mucunoides</i>	1.51 ^b	0.71 ^a	Small, round, black	Spread

Different superscript letters in the same column show significant difference ($P < 0.05$). Centre position means the noodles were only observed in the primary roots.

Table 5. The weight of legume and grass roots

Kind of legumes	Weight of legume roots (g)		Weight of grass roots (g)
	Legume areas	Grass areas	
<i>G. sepium</i>	81.2 ^a	9.0 ^c	135.0 ^a
<i>L. leucocephala</i>	60.0 ^b	27.5 ^b	150.5 ^a
<i>C. calothyrsus</i>	14.0 ^c	2.8 ^d	34.6 ^c
<i>C. mucunoides</i>	15.5 ^c	35.0 ^a	60.0 ^b

Different superscript letters in the same column show significant difference ($P < 0.05$).

biological condition of the plants and then affects the plant growth and production. Walzi et al. (2012) showed that more legume roots enhance nutrient distribution for their companion. Surprisingly our data showed that the highest weight of legume roots in the legume area and quite low in grass area observed in *G. sepium* produced highest weight of grass root. In the opposite observed in *C. mucunoides* which had high legume root weight in legume area produced low weight of grass root. The weight root or structural of roots influence the grass production will be discussed later in the production paragraph.

Grass heights and shoots

The grass height was significantly influenced by kind of legumes intercropped, while number of shoots in clumps was not significantly influenced by the treatments (Table 6). The number of shoots was not significantly influenced even though in control, the one was not intercropped which had the highest number, 30 % than the lowest. The highest *P. maximum* was observed in the one intercropped with *G. sepium* (160.7 cm) followed with *C. calothyrsus* (140.0 cm). Grass that grew without any legume integrated showed lowest height (98.3 cm). The same result has been reported by Sajimin et al. (2005) and Sajimin & Jarmani (2014) that the height of *P. maximum* is 118.2 cm/clump in

monocultures, while intercropping with *Clitoria ternatea* it reaches 156.0 cm/clump. Intercropping with legume roots produced more grass due to the transportation of nutrients from legume areas. The same result was also reported by Ojo et al. (2013) that *P. maximum* grown faster if intercropped with a legume of *Lablab purpureus* compared with the one without. Onyeonogu & Asiegbe (2013) the highest tiller number per meter square was obtained in *P. maximum* intercropping with the legumes *Stylosanthes hamata*.

Forage production

Legume forage production was significantly influenced by kind of legumes (Table 7). The highest forage production of the legumes was observed in *C. mucunoides* followed by the others. This plant is a shrub legume, while others are tree legumes. This experiment was more to see the grass production, however, since the legumes also share the forage production. The legume production should be noticed. Each legume species significantly produced different forage amounts due to the difference in morphology and genetics. This legume had the highest root weight in legume area (Table 4) which may take part in using the nutritional elements from the grass soil for the legume growth resulted the reduction of grass production.

Table 6. Heights and shoot numbers per clumps of grass grown integrated with legumes

Kind of legumes	Height of grass (cm)	Number of shoots per clumps
<i>G. sepium</i>	160.7 ^a	9.8
<i>L. leucocephala</i>	135.5 ^b	12.3
<i>C. calothyrsus</i>	140.0 ^{ab}	9.5
<i>C. mucunoides</i>	125.5 ^b	10.8
Control	98.3 ^c	15.0

Different superscript letters in the same column show significant difference (P<0.05).

Table 7. Forage production of legumes and *P. maximum* cv Riversdale intercropped with the legume roots

Kind of legumes	Forage production (g DM/pot)	
	Legumes	<i>P. maximum</i>
<i>G. sepium</i>	21.0 ^c	54.7 ^a
<i>L. leucocephala</i>	21.1 ^c	50.7 ^{ab}
<i>C. calothyrsus</i>	25.4 ^{bc}	47.4 ^{ab}
<i>C. mucunoides</i>	51.9 ^a	30.0 ^b
Control	No plants	40.7 ^{ab}

Different superscript letters in the same column show significant difference (P<0.05).

Except for *C. mucunoides* the productions of grass intercropped with legumes were significantly higher than the control. The highest production of grass was observed at the one intercropped with *G. sepium* (54.7 g/plant) followed by the one with *L. leucocephala* (50.7 g), *Caliandra* (47.4 g), while the one with *C. mucunoides* had the lowest (30.0 g) (Table 7). The production of *L. leucocephala* was not significantly different to those of *G. sepium* and *C. calothyrsus*. The lowest grass production in the one with *C. mucunoides* might be related with short grass due to limited nutrient caused by poor quality of legume root nodules (Table 4 and 6), and the least weight of grass root. The intercropping system was not affected for the grass production, however it produced the highest legume forage.

All the effected intercroppings for grass productions were from tree legumes, those were *G. sepium*, *L. leucocephala*, and *C. calothyrsus*. The leaves of the height plants might drop to grass area and result the increase of nutritional elements in the grass soil, opposite to *C. mucunoides* which was quite short and only dropped its leaves in legume areas. All tree legumes gave higher P translocation higher than the negative control (Table 1). In the top of that the *Rhizobium* in their nodules which have the ability to fix the nitrogen will take part in improving the growth. The same results those grass biomass productions were increased by legume intercropping have been reported by Baba et al. (2011) and Abdullah et al. (2014). Darmadeh (2013) also reports that integration of peanut increases the biomass production of maize.

Intercropping with *G. sepium* gave the highest production, since it translocated the highest P to the soil grass, produced more effective root nodules, as well as the tallest grass and heighest grass root weight. Although the grass production was the highest, the legume production was quite low. Therefore, the total amount of the forage production from grass and legume was comparable with the one intercropped with *C. mucunoides*. This experiment was carried out in the pot or in the limited area. However, it already showed the positive effect of legume intercropping. Scaling up should be evaluated in the farm and the evaluation also should consider the forage production of the legumes.

CONCLUSION

It can be concluded from the experiment that intercropping *Panicum* grass with the legume roots increase P availability in the grass area. The P translocation was proved by the detection of ³²P radio isotop injected in the legume area. The increase of the P in the grass area and the activity of *Rhizobium* in nitrogen fixation enhanced the production of the grass.

The best grass production was observed when the grass was intercropped with *G. sepium*.

REFERENCES

- Abdullah AA, Karti PDMH, Chozin MA, Astuti DA. 2014. Evaluasi produktivitas dan daya saing dari *Brachiaria decumbens*, *Centrosema pubescens* dan *Clitoria ternatea* sebagai tanaman tunggal dan campuran pola pemotongan di lahan gambut. JITV. 19:81-90.
- Anantawiroon P, Tudsri S, Ishit Y. 2006. The effect of intercropping with four tropical legume species on the yield and quality of Napier grass in Thailand. Kasetsart J Nat Sci. 40:616-624.
- Alan E. Richardson AE, Barea JM, McNeill AM, Prigent-Combaret J. 2009. Acquisition of phosphorus and nitrogen in the rhizosphere and plant growth promotion by microorganisms. Plant Soil. 321:305-339. doi: 10.1007/s11104-009-9895-2.
- Agus F, Wiratno, Suwardi. 2015. Status of Indonesian soil resources. Asian soil partnership consultation workshop on sustainable management and protection of soil resources. Bangkok (Thailand): Food and Agriculture Organization.
- Baba M, Halim RA, Alimon AR, Abubakar I. 2011. Grass-legume mixture for enhanced forage production: Analysis of dry matter yield and competition indices. Afr J Agric Res. 6:5242-5250.
- Belel MD, Halim RA, Raffi MY, Saud HM. 2014. Intercropping of corn with some selected legumes for improved forage production: A review. J Agric Sci. 6:48-62.
- Catchpoole DW. 1988. The contribution of tree legumes to the nitrogen economy and forage production in the humid tropic (Thesis). [(New South Wales) Australia]: University of New England.
- Darmadeh M. 2013. Intercropping two varieties of maize (*Zea mays* L) and peanut (*Arachis hypogea* L): Biomass yield and intercropping advantages. Int J Agric Forest. 3:7-11.
- Exner DN, Davison DG, Ghaffarzadeh M, Cruse R M 1999. Yields and returns from strip intercropping on six Iowa farms. Am J Alter Agric. 14:69-77.
- Elgersma A, Schleggers H, Nassiri M. 2000 Interactions between perennial ryegrass (*Lolium perenne* L.) and white clover (*Trifolium repens* L.) under contrasting N availability: Productivity, seasonal patterns of species composition, N₂ fixation, N transfer and N recovery. Plant Soil. 221:281-299.
- Eskandori H, Ghanbari A, Javonmard A. 2009. Intercropping of cereals and legumes for forage production. Nat Sci Biol. 1:7-13.
- Fustec J, Lesuffleur F, Mahieu S, Cliquet JB. 2010. Nitrogen rhizodeposition of legumes: A review. Agron J Sustain Develop. 30:57-66. INRA EDP Sci. doi: 10.1051/agro/2009003.

- Franzini VI, Muraoka T, Mendes FL. 2009. Ratio and rate effects of ^{32}P -triple super phosphate and phosphate rock mixtures on corn growth. *Sci Agric Piracicaba Braz.* p. 71-76.
- Guo F, Yost RS, Hue NV, Evensen CI, Silva JA. 2000. Changes in phosphorus fractions in soils under intensive plant growth. *Soil Sci Am. J.* 64:1681-1689.
- Gomez KA, Gomez AA. 1984. *Statistical procedures for agricultural research*. 2nd ed. IRRI. John Wiley and Sons, Inc.
- Hakim N, Nyakpa NY, Lubis AM, Nugroho SG, Saul ML, Diha MA, Go BH, Bailey HH. 1986. *Dasar-dasar ilmu tanah*. BKS. PTN/USAID. Unila Lampung.
- Horst WJ, Eticha D, Kamh M, Wang Y, Stival Da Silva AL, Stass A. 2006. Identification and characterization of aluminium resistant, phosphorus-efficient plant genotypes adapted to tropical acid soils. Hannover (Germany): Institute for Plant Nutrition, University of Hannover, Germany. Proceeding series. Management practices for improving sustainable crop production in tropical acid soil. IAEA.
- Karama AS, Marzuki AR, Manwan I. 1991. Penggunaan pupuk organik pada tanaman pangan. *Prosiding Lokakarya Nasional Efisiensi Penggunaan Pupuk V*. Bogor (Indones): Indonesian Center for Animal Research and Development. p. 395-485.
- Kalaivanan D, Sudhir K, Kotur SC. 2014. Effect of different levels and placement of ^{32}P labelled single super phosphate on absorption of phosphorus by *Banana* cv. *robusta* under high density planting. *Vegetos.* 27:68-75.
- Latati M, Balvet D, Alkama N, Laoufi H, Drevon JJ, Gerard F, Pansu M, Ounane SM. 2014. The intercropping cowpea-maize improves soil phosphorus availability and maize yields in an alkaline soil. *Plant Soil.* 385:181-191.
- Li L, Zhang FS, Li XL, Christie P, Sun JH, Yang SC, Tang CX. 2003 Interspecific facilitation of nutrient uptake by intercropped maize and fababean. *Nutr Cycl Agroeco.* 65:61-71.
- Mulyani A, Hikmatullah, Subagyo H. 2004. Karakteristik dan potensi tanah masam lahan kering di Indonesia. *Prosiding Simposium Nasional Pendugaan Tanah Masam*. Bogor (Indones): Pusat Penelitian dan Pengembangan Tanah dan Agroklimat. p. 1-32.
- Mutegi JK, Mugendi DN, Verchot LV, Kung'u JB. 2008. Combining Napier grass with leguminous shrubs in contour hedgerows controls soil erosion without competing with crops. *Agroforest Syst Springer Sci.* 13 p.
- Ojo VOA, Dele PA, Amole TA, Anele UY, Adeoye SA, Hassan OA, Olanite JA, Idowu OJ. 2013. Effect of intercropping *Panicum maximum* Var. Ntchisi and *Lablab purpureum* on the growth, herbage yield and chemical composition of *Panicum maximum* var. Ntchisi at different harvesting times. *Pak J Biol Sci.* 16:1605-1608.
- Richardson AE, Barea JM, McNeill AM, Prigent-Combaret C. 2009. Acquisition of phosphorus and nitrogen in the rhizosphere and plant growth promotion by microorganisms. *Plant Soil.* 321:305-339. doi: 10.1007/s11104-009-9895-2.
- Sajimin, Jarmani SN. 2014. Pengaruh interval panen terhadap produksi kualitas rumput *P. maximum* cv Purple Guinea sebagai pakan ternak di Kabupaten Blora. Kaiin EM, Tappa B, Widyastuti Y, Said S, Agung PP, editors. *Prosiding Seminar Nasional Peran Bioteknologi dalam Peningkatan Populasi dan Mutu Genetik Ternak Mendukung Kemandirian Daging dan Susu Nasional*. Bogor (Indones): Lembaga Ilmu Pengetahuan Indonesia. p. 110-117.
- Onyeonogu CC, Asiegbu JE. 2013. Harvest frequency effect on plant height, grass tiller production, plant cover and percentage dry matter production of some forage grass and legumes in the derived Savannah Nigeria. *Afr J Agr Res.* 8:608-618.
- Sajimin, Sutedi E, Purwantari ND, Prawiradiputra BR, 2005. Agronomi rumput benggala (*Panicum Maximum* Jacq) dan pemanfaatannya sebagai rumput potong. Subandriyo, Diwyanto K, Inounu I, Prawiradiputra BR, Setiadi B, Nurhayati, Priyanti A, editors. *Prosiding Lokakarya Nasional Tanaman Pakan Ternak*. Bogor (Indones): Pusat Penelitian dan Pengembangan Peternakan. p. 121-129.
- Saidu A, Abayomi YA. 2015. Interactive effects of organic and inorganic fertilizers on the performance of Upland Rice (*Oryza sativa* l.) Cultivars. *Int J Agr Sci.* 5:399-406.
- Walzi KP, Rasmussen J, Jensen HH, Eriksen J, Soegard K, Rasmussen J. 2012. Nitrogen transfer from forage legumes to nine neighbouring plants in a multi-species grassland. *Plant Soil.* 350:71-84.
- Xiao Y, Li L, Zhang F. 2004. Effect of root contact on interspecific competition and N transfer between wheat and fabean using direct and indirect ^{15}N techniques. *Plant Soils.* 262:45-54.